

MSc Quantitative Life Science

Study Program Handbook



Subject-specific Examination Regulations for Quantitative Life Science (QLS)

The subject-specific examination regulations for Quantitative Life Science are defined by this program handbook and are valid only in combination with the General Examination Regulations for Master degree programs (“General Master Policies”).

Upon graduation students in this program will receive a Master of Science (MSc) degree with a scope of 120 ECTS credit points (CP) (for specifics see chapter 3 of this handbook).

Valid for all students starting their studies in Fall 2025

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1 Program Overview

1.1 Background

Life Science is the study of living organisms and the processes of life. It embraces several classical scientific disciplines including aspects of biology, biochemistry, medicine, pharmacy, chemistry and bioengineering, and it thus contributes to solving many societal challenges ranging from health care and nutrition to global warming. While traditionally considered a science based on qualitative experiments, the emergence of new technologies, like next-generation sequencing, super-resolution imaging and high-resolution mass spectrometry, lead to a wealth of quantitative data offering the opportunity for a deeper understanding of the most fundamental processes of life.

According to Galileo “Mathematics is the language, in which God has written the Universe” or in the words of S. Devi “Mathematics is a systematic effort of solving puzzles posed by nature”. Quantitative Life Science relates biological systems to mathematics. In the last century, living organisms were still considered too complex for quantitative assessment and computational modelling due to the plethora of different molecules and processes that govern cellular biology and interaction. Additionally, any living organism is exposed to a multitude of environmental factors and xenobiotics that further influence organismal function and behavior.

In the 21st century, however, the situation has dramatically changed. Modern experimental technologies emerged that allow a comprehensive analysis of living organisms, which have been termed the science of Omics. For example, modern mass spectrometry can obtain information on the molecular composition and content of a biological sample within a few hours of measurement time. Likewise, the genomes of whole organisms containing genetic information in billions of nucleotides can nowadays be sequenced within hours or days, thereby generating terabytes of data a month in many laboratories. Concomitantly, modern data science has caught up in the last decade with this development, providing suitable hardware to store and process the immense amount of data obtained. With the help of novel approaches such as machine learning and artificial intelligence, scientists are now able to not only manage big datasets but also extract meaningful insights and ultimately knowledge from biological databases. All these developments call for a Master program reflecting these developments in the Life Sciences offered by Constructor University.

To leverage this vast amount of data and perform successful research in this age of big data, scientists thus require logical rigor and numerical literacy allowing them to analyze and interpret these new forms of data. Hereby the current developments in the field of information technology, like *high-performance computing* and machine learning, will prove invaluable tools towards these aims. Moreover, developing new hypothesis and design experiments exploiting the full potential of these new technologies requires a deep understanding of their scopes and limitations.

While mastering these skills is crucial to participate in this on-going revolution in the field of life science and assume leadership roles in scientific research in academia and industry. Therefore, and in contrast to *classical* Life Science, Quantitative Life Science places an emphasis on numerical data, obtained by experimental measurements, processed and generalized by mathematics and the tools of modern data science.

1.2 Concept

The Quantitative Life Science (QLS) Master Program at Constructor University prepares Life Science students for a successful career in academia or industry through an interdisciplinary education combining cutting-edge quantitative experimental techniques as well as the computational expertise to generate, critically evaluate, fully understand and utilize the associated data. Additional cornerstones of

the program are a strong research component, specialization through diverse electives, and transferable skill courses reflecting real-life science and business requirements.

The majority of Life Science graduates continue to study for a PhD degree. Hence, our study program MSc Quantitative Life Science provides the opportunity for individual specialisation as well as the skills and knowledge to successfully conduct independent research in a PhD program at Constructor University or other world-class universities. An international campus and a close interaction between teachers and students provide an intellectual and social environment that offers numerous opportunities for personal development and for working in diverse teams.

Students will also learn how to operate as a member of a team in a research group and contribute as a reliable and valuable collaborator, and even a responsible leader in larger projects. The faculty advises, encourages, and supports students to think and study independently, conduct autonomous background reading, solve problems alone or in teams, and bring new ideas and solutions to seminars and tutorials for discussion. The MSc in Quantitative Life Science is a truly interdisciplinary program. It benefits from a broad faculty expertise offering modules that range from various branches of Life Science and Chemistry interlinked with computer science and software engineering.

1.3 Qualification Aims

1.3.1 Educational Aims

The MSc in Quantitative Life Science aims to educate independent scientific researchers able to pursue a successful career at the intersection of life science and computer science.

For this, the students will broaden and deepen their knowledge of innovative computational approaches, state-of-the-art experimental techniques, and theoretical concepts in Quantitative Life Science with a particular focus on the Omics disciplines. These technical and programming skills combined with subject-matter expertise will enable students to work in a scientific environment.

Students will gain experience, applying these skills in actual research projects and learn how to advance scientific knowledge by developing original research hypotheses and designing, planning, and executing the wet-lab and dry-lab experiments to test them. They will also gain the necessary numerical literacy and scientific rigor to scrutinize, interpret and defend experimental results or computational simulations to international scientific standards.

Students will refine their skills to visualize their data, efficiently communicate results and concepts to scientific audiences through oral presentations, written research proposals or scientific publications, and discuss them with their peers.

Throughout the MSc program the students will refine their ability to reflect on their own performance, work in diverse teams and act as a responsible, ethical, and effective stakeholder in the scientific community and beyond.

1.3.2 Intended Learning Outcomes

By the end of this program, students will be able to:

1. Describe complex technical and conceptional advances in the Life Sciences in depth and with specific focus on Omics technologies, like genomics, transcriptomics, proteomics, and metabolomics.
2. Understand common computational tools, algorithms, and databases for data processing, analysis and visualization used in the life sciences.

3. Critically examine and creatively apply intricate techniques to perform quantitative experiments, obtain reliable data, and answer complex scientific questions in the life sciences at high scientific and ethical standards.
4. Scrutinize and implement innovative bioinformatics approaches to obtain, examine, and compare Life Science data.
5. Construct comprehensive mathematical models and detailed computational simulations to generate and test hypothesis in the field of biology or chemistry.
6. Scrutinize and critique scientific results, experimental approaches, and research hypothesis from all fields of quantitative life science.
7. Construct original and innovative research hypothesis and design experiments to test them individually or as a team in actual research projects and a master thesis.
8. Apply professional writing, communication, and presentation techniques, combined with state-of-the-art visualization tools to effectively communicate with scientific and non-scientific audiences.
9. Understand the chances, risks, and approaches of applying scientific research in modern society considering ethical and commercial aspects while following and defending ethical, scientific, and professional standards.
10. Take responsibility for their own learning, personal and professional development and role in diverse teams, evaluating critical feedback and self-analysis.

1.4 Target Audience

The study program MSc Quantitative Life Science is designed for students with a strong background in Life Science and related subjects, aiming to pursue a career in research and are excited to exploit the potential of big-data technologies and computer-assisted analysis.

As a consecutive second-degree program, the students should hold a BSc in related fields such as biology, cell biology, biochemistry, chemical biology, chemistry, pharmacy, biotechnology, and bioinformatics, and be proficient in English, the language of education.

Moreover, the program aspires to attract international students from a variety of cultural and geographical backgrounds to foster a diverse community with room for exchange and creativity.

1.5 Career Options

The program primarily prepares the students for a career as independent scientists either in academia or industry. Staying in academia, we expect the students to pursue a PhD degree, and the independent research in the master thesis as well as advanced electives, provide the perfect basis for this endeavour. In industry, graduate student can, thanks to their technical skills, directly contribute as research scientists, with the *transferable skills courses* providing the necessary skills to assume leadership roles.

Future graduates of this program will transfer to PhD programs or will take up positions in various fields and industries, most notably the biotechnology sector, pharmaceutical industry, agrochemical industry, analytical service providers, food industry NGOs, academia or governmental regulatory authorities.

Furthermore, the logical thinking, numerical literacy and computational skills transferred in the program also allows the graduate to pursue careers in highly quantitative Jobs, like management consulting or financial services.

The Career Service Center (CSC) helps students in their career development. It provides students with high-quality training and coaching in CV creation, cover letter formulation, interview preparation, effective presenting, business etiquette, and employer research as well as in many other aspects, thus helping students identify and follow up on rewarding careers after graduating from Constructor University. For further information, please contact the Career Service Center (CSC) (<https://constructor.university/student-life/career-services>). Furthermore, the Alumni Office helps students establish a long-lasting and global network which is useful when exploring job options in academia, industry, and elsewhere.

1.6 Admission Requirements

The Quantitative Life Science master program requires students to hold at least a good bachelor's degree in the areas of Life Science or related Areas. Applicants need to prove a strong interest in the contents of the study program in a motivation letter. The general "Admission and Enrollment Policies" of Constructor University apply (see [Academic policies | Constructor University](#)). Social commitment as well as extracurricular and voluntary activities during undergraduate studies, e.g. university service, clubs, varsity, social work, etc. will be considered.

Additionally, participants should possess elevated analytical, problem solving and verbal communication skills which must be substantiated in recommendation letters.

Study at Constructor University takes place in a highly intercultural environment. It is therefore necessary to be willing to join such a multicultural-international community and work together with students and faculty across various fields of interest at Constructor University.

Admission to Constructor University is selective and based on a candidate's university achievements, recommendations and self-presentation. Students admitted to Constructor University demonstrate exceptional academic achievements, intellectual creativity, and the desire and motivation to make a difference in the world.

The following documents need to be submitted with the application:

- Letter of motivation
- Curriculum vitae (CV)
- Official or certified copies of university transcripts
- Bachelor's degree certificate or equivalent
- Language proficiency test results (minimum score of 90 (TOEFL), 6.5 (IELTS) or 110(Duolingo)).
- Copy of Passport
- Letter of recommendation

Formal admission requirements are subject to higher education law and are outlined in the Admission and Enrollment Policy of Constructor University.

For more detailed information about the admission visit:

[Application Information | Constructor University](#)

1.7 More information and contacts

For more information on the study program please contact the Study Program Coordinator:

- 1) Dr Nikolai Kuhnert, Professor of Chemistry, nkuhnert@constructor.university
- 2) Dr Felix Jonas, Assistant Professor of Biochemistry, fjonas@constructor.university

or visit our program website: <https://constructor.university/programs/graduate-education/quantitative-life-science>

For more information on Student Services please visit:
<https://constructor.university/student-life/student-services>

2 The Curriculum

2.1 The Curriculum at a Glance

The MSc in Quantitative Life Science is a 2-year full-time in-presence program with 120 ECTS points, divided into four semesters. The first three semesters are each composed of a mixture of core, elective, transferable skills courses, and a strong research component, while the fourth and final semester is dedicated to the Master thesis.

The first semester is a foundational semester, in which students from different educational backgrounds primarily get acquainted with the most foundational concepts in the Life Sciences and acquire solid computational skills. The second and third semester focus on different aspects of the Quantitative Life Science and further allow the students to explore their own interests in research projects and electives, while also refining their transferable skills. The fourth and final semester is reserved for the Master Thesis where students can apply their skills in their own independent research project.

The modules are grouped in five categories, the mandatory CORE modules (30 CP), the mandatory elective modules (15 CP), the transferable skill modules (15 CP), the research modules (30 CP) and Master Thesis (30 CP).

2.2 Schematic Study Scheme

MSc Degree in <i>Quantitative Life Science (QLS)</i> (120CP)					
Semester					
4	Master Thesis (m, 30 CP)				
3	Computational Life Science - Modelling and Simulations (m, 5 CP)	Omics II - Proteomics and Metabolomics (m, 5 CP)	Lab Rotation QLS III (m, 10 CP)	Intellectual Property and Commercialization (m, 5 CP)	Elective III (me, 5 CP)
2	Experimental Techniques (m, 5 CP)	Omics I - Genomics and Transcriptomics (m, 5 CP)	Lab Rotation QLS II (m, 10 CP)	Scientific Writing (m, 5 CP)	Elective II (me, 5 CP)
1	Guided Self-study (Chemistry, Biology, Math/Physics) (m, 5 CP)	Data tools for the Life Sciences (m, 5 CP)	Lab Rotation QLS I (m, 10 CP)	Scientific Presentations (m, 5 CP)	Elective I (me, 5 CP)
Area	CORE 30 CP		Research 30 CP	Transferable Skills 15 CP	Elective 15 CP

m = mandatory

me = mandatory elective

MSc Degree in *Quantitative Life Science*
Elective Modules

Quantitative Life Science Electives			
	Molecular Simulations (me, 5 CP)	Biochemical Engineering - From Cells to Processes (me, 5 CP)	Structure Elucidation (me, 5 CP)
	Machine Learning in Cheminformatics (me, 5 CP)	Quantitative Cell Biology (me, 5 CP)	Supramolecular Chemistry (me, 5 CP)
Area	Computational LS	Cellular LS	Chemical LS

2.3 Study and Examination Plan

MSc Degree in Quantitative Life Science							
Matriculation Fall 2025							
Module Code	Program-Specific Modules	Type	Assessment	Period ¹	Status ²	Semester	CP
Semester 1							30
CORE Area							10
MQLS-CO-01	Module: Guided Self-Study				m	1	5
MQLS-CO-01	Guided Self-Study	Self-study/Tutorial	Written examination	Examination period			
MQLS-CO-02	Module: Data Tools for the Life Sciences				m	1	5
MQLS-CO-02-A	Data Tools for the Life Sciences Lecture	Lecture/Tutorial	Written examination	Examination period			2.5
MQLS-CO-02-B	Data Tools for the Life Sciences Project	Project	Project report	During semester			2.5
Elective Area							5
- students choose one module from those listed below							
Research Area							10
MQLS-RES-01	Module: Lab Rotation QLS I				m	1	10
MQLS-RES-01-A	Lab Rotation QLS I - Lab	Lab	Laboratory Report	Examination period			7.5
MQLS-RES-01-B	Lab Rotation QLS I - Seminar	Seminar	Presentation	During semester			2.5
Transferrable skills Area							5
MQLS-TS-01	Module: Scientific Presentations				m	1	
MQLS-TS-01	Scientific Presentations	Lecture/Seminar	Presentation	During semester			
Semester 2							30
CORE Area							10
MQLS-CO-03	Module: Experimental Techniques				m	2	5
MQLS-CO-03	Experimental Techniques	Lecture	Written examination	Examination period			
MQLS-CO-04	Module: Omics I - Genomics and Transcriptomics				m	2	5
MQLS-CO-04	Omics I - Genomics and Transcriptomics	Lecture	Written examination	Examination period			
Elective Area							5
- Students choose a module from those listed below.							
Research Area							10
MQLS-RES-02	Module: Lab Rotation QLS II				m	2	10
MQLS-RES-02-A	Lab Rotation QLS II - Lab	Lab	Laboratory Report	Examination period			7.5
MQLS-RES-02-B	Lab Rotation QLS II - Seminar	Seminar	Presentation	During semester			2.5
Transferrable Skills Area							5
MQLS-TS-02	Module: Scientific Writing				m	2	5
MQLS-TS-02	Scientific Writing	Seminar	Term Paper	During semester			
Semester 3							30
CORE Area							10
MQLS-CO-06	Module: Computational Life Sciences - Modelling and Simulations				m	3	5
MQLS-CO-06	Computational Life Sciences - Modelling and Simulations	Lecture	Written examination	Examination period			
MQLS-CO-05	Module: Omics II - Microbiomics, Proteomics, and Metabolomics				m	3	5
MQLS-CO-05	Omics II - Microbiomics, Proteomics, and Metabolomics	Lecture	Written examination	Examination period			
Elective Area							5
- Students choose a module from those listed below.							
Research Area							10
MQLS-RES-03	Module: Lab Rotation QLS III				m	3	10
MQLS-RES-03-A	Lab Rotation QLS III - Lab	Lab	Laboratory Report	Examination period			7.5
MQLS-RES-03-B	Lab Rotation QLS III - Seminar	Seminar	Presentation	During semester			2.5
Transferrable Skills Area							5
MQLS-TS-03	Module: Intellectual Property and Commercialization				m	3	5
MQLS-TS-03	Intellectual Property and Commercialization	Lecture	Oral Examination	flexible			

Semester 4					30
Master Thesis					30
MQLS-THE-01	Module: Master Thesis QLS		m	4	30
MQLS-THE-01	Master Thesis QLS		Thesis		
			Oral Examination		
Total CP					120

¹ Each lecture period lasts 14 semester weeks and is followed by reading and examination days. Written examinations are centrally scheduled during weeks 15 and 16. For all other assessment types, the timeframes indicated in the above

² m = mandatory, me = mandatory elective

Elective Area					
<i>Students choose 15 CP of mandatory electives</i>					
Computational Life Science					10
MQLS-COM-02	Module: Machine Learning in Cheminformatics		me	3	5
MQLS-COM-02	Machine Learning in Cheminformatics	Lecture, Seminar	Project Assessment	During semester	
MQLS-COM-01	Module: Molecular Simulations		me	3	5
MQLS-COM-01-A	Molecular Simulations Lecture	Lecture	Written examination	Examination period	2.5
MQLS-COM-01-B	Molecular Simulations Lab	Lab	Laboratory report	During semester	2.5
Chemical Life Science					10
MQLS-CHE-01	Module: Supramolecular Chemistry		me	1 or 3	5
MQLS-CHE-01	Supramolecular Chemistry	Lecture	Written examination	Examination period	
MQLS-CHE-02	Module: Structure Elucidation		me	2	5
MQLS-CHE-02	Structure Elucidation	Lecture	Written examination	Examination period	
Cellular Life Science					10
MQLS-CEL-01	Module: Quantitative Cell Biology		me	4	5
MQLS-CEL-01-A	Quantitative Cell Biology seminar	Seminar	Written examination	Examination period	2.5
MQLS-CEL-01-B	Quantitative Cell Biology tutorial	Tutorial	Presentation	During semester	2.5
MQLS-CEL-02	Module: Biochemical Engineering - From Cells to Processes		me	1 or 3	5
MQLS-CEL-02-A	Microbial Engineering	Lecture			2.5
MQLS-CEL-02-B	Advanced Bioprocess Engineering and Digitalization	Lecture	Written examination	Examination period	2.5
Other Areas					
<i>- A set of modules from other study programs will be provided as further electives. Please see CampusNet for further information</i>					
Total CP					30

2.4 Core Area: 30 CP

Modules of the CORE area convey the essential concepts as well as key experimental and computational approaches in quantitative life science. Hence all of the following six Core modules (30 CP) are mandatory (m) to pursue a master's degree in quantitative Life Science:

- CORE Module: Guided self-study (m, 5 CP)
- CORE Module: Data Tools for the Life Sciences (m, 5 CP)
- CORE Module: Experimental Techniques (m, 5 CP)
- CORE Module: Omics I - Genomics and Transcriptomics (m, 5 CP)
- CORE Module: Omics II - Microbiomics, Proteomics, and Metabolomics (m, 5 CP)
- CORE Module: Computational Life Sciences - Modelling and Simulations (m, 5 CP)

2.5 Research Area: 30 CP

The QLS master program provides the unique opportunity conduct actual research to explore different scientific areas by providing three laboratory rotations (semesters 1-3, 10 CP per semester). To pursue a master's degree in quantitative Life Science, the following three Research modules (30 CP) need to be taken as mandatory modules (m):

- Research: Lab Rotation QLS I (m, 10 CP)
- Research: Lab Rotation QLS II (m, 10 CP)
- Research: Lab Rotation QLS III (m, 10 CP)

2.6 Elective Area: 15 CP

To further deepen their knowledge in selected areas, students may choose elective modules. To fulfil the requirement for a QLS master, students need to take three of the following mandatory elective modules (me). To meet the credit requirement, any modules from the three indicated areas can be combined.:

Computational Life Science:

- CHOICE Module: Machine Learning in Cheminformatics (me, 5 CP)
- CHOICE Module: Molecular Simulations (me, 5 CP)

Chemical Life Science:

- CHOICE Module: Supramolecular Chemistry (me, 5 CP)
- CHOICE Module: Structure Elucidation (me, 5 CP)

Cellular Life Science:

- CHOICE Module: Quantitative Cell Biology (me, 5 CP)
- CHOICE Module: Biochemical Engineering - From Cells to Processes (me, 5 CP)

2.7 Transferable Skills Area: 15 CP

In the transferable skills area, three mandatory modules (15 CP) need to be taken. In these modules, the students learn how to present scientific data in oral and written format. Furthermore, they will explore options to secure their intellectual property rights in a scientific setting and for commercialization.

- Transferable Skills: Scientific Presentations (m, 5 CP)
- Transferable Skills: Scientific Writing (m, 5 CP)
- Transferable Skills: Intellectual Property and Commercialization (m, 5 CP)

2.8 Master Thesis (30 CP)

In the fourth semester, students conduct independent research in one of the participating research groups. To complete this module, students write a mandatory master thesis guided and supported by their research supervisor, worth of 30 credit points.

- Thesis Module: Master Thesis QLS (m, 30 CP)

3 Quantitative Life Science Graduate Program Regulations

3.1 Scope of these Regulations

The regulations in this handbook are valid for all students who entered the Quantitative Life Science graduate program at Constructor University in Fall 2025. In case of conflict between the regulations in this handbook and the general policies for Master Studies, the latter apply (see [Academic policies | Constructor University](#)).

In exceptional cases, certain necessary deviations from the regulations of this study handbook might occur during the course of study (e.g., change of the semester sequence, assessment type, or the teaching mode of courses).

Updates to Study Program Handbooks are based on the policies approved by the Academic Senate on substantial and nonsubstantial changes to study programs. Students are integrated in the decision-making process through their respective committee representatives. All students affected by the changes will be properly informed.

In general, Constructor University therefore reserves the right to change or modify the regulations of the program handbook also after its publication at any time and in its sole discretion.

3.2 Degree

Upon successful completion of the study program, students are awarded a Master of Science (MSc) degree in Quantitative Life Science.

3.3 Graduation Requirements

In order to graduate, students need to obtain 120 credit points. In addition, the following graduation requirements apply:

- Students need to complete all mandatory components of the program as indicated in chapter 2 of this handbook

3.4 Other Program-specific Policies & Practices

Close contact and cooperation between program representatives and students are crucial. Therefore, regular meetings are held to continuously evaluate the program, its modules and workshops, supervision, and opportunities. In doing so, the Study Program Chair and involved faculty gain important insights into students' experiences, demands, and overall impressions of the program. On the module component level, students are asked to perform module component evaluations to ensure that the modules are high-quality and that lecturers can make any necessary changes.

The study program chair makes intensive use of this feedback as well as feedback from laboratory heads to improve the learning environment, the program's offering, and its progress. The current program was shaped through input from previous experiences and discussions with several stakeholders, including students and industry practitioners.

In exceptional cases, certain necessary deviations from the regulations of this study handbook might occur during the course of study (e.g., change of the semester sequence, assessment type, or the teaching mode of courses). Constructor University Bremen reserves therefore the right to modify the regulations of the program handbook.

For biological and chemical safety in laboratory work both regulations within German law (e.g. Arbeitsschutzgesetz, Gefahrstoffverordnung, Chemikaliengesetz etc) apply as well as University internal health and safety regulations. Students are handed out the relevant regulations at the beginning of the program. Furthermore, attendance of S1, S2, and Laser safety instructions are mandatory prior to any laboratory work. Before joining a research group, students are given laboratory specific safety instructions (research group specific laboratory regulations).

4.1 Biochemical Engineering - From Cells to Processes

Module Name	Biochemical Engineering - From Cells to Processes
Module Code	2025-MQLS-CEL-02
Module ECTS	5
Study Semester	Mandatory status for: None Mandatory Elective status for: - 2025-QLS-MSc 1
Duration	1 Semester
Program Affiliation	2025-QLS-MSc (Quantitative Life Science)
Module Coordinator(s)	Prof. Dr. Elke Nevoigt Prof. Dr. Katrin Rosenthal

Forms of Learning and Teaching	
Lecture	35
Exam Preparation	35
Independent Study	55
Workload Hours	125 hours

Module Components	Number	Type	CP
Microbial Engineering	MQLS-CEL-02-A	Lecture	2.5
Advanced Bioprocess Engineering and Digitalization	MQLS-CEL-02-B	Lecture	2.5

Module Description

Many valuable products such as enzymes, pharmaceutical proteins, small-molecule pharmaceuticals, biofuels, commodities and fine chemicals can be produced from renewable raw materials using living cells or their ingredients. This course combines strategies on how to design and optimize bioprocesses by engineering either i) the cellular factory or ii) the reaction. With regard to cell factory engineering, cutting-edge methods of synthetic biology that support metabolic engineering will be the main focus.

The second module component explores the fundamental concepts and distinctions between in vitro and in vivo biotransformations, emphasizing their implications and applications in bioprocessing. Students will analyze and evaluate integrated process

development approaches for both upstream and downstream operations, gaining insights into the optimization of bioprocess efficiency through digital twins. Additionally, the module covers the application of computational tools and digitalization techniques, including computer-aided design software to enhance bioreactor and bioprocess design.

Recommended Knowledge

- Previous exposure to the specified course material is helpful but not essential.

- Fundamentals of Molecular Biology and

Genetic Engineering, Microbiology, Metabolic

Pathways, Enzymes

Usability and Relationship to other Modules

This module is for students who want to extend their studies in Metabolic

Engineering, Biotechnology, and Biochemical Engineering.

Intended Learning Outcomes

No	Competence	ILO
1	Examine	Examine how genetic engineering and synthetic biology has changed our opportunities to create microbial cells that can be exploited for the production of a target molecule
2	Understand	Understand the necessity of the 'design-build-test-learn cycles' in targeted microbial strain development
3	Discuss	Discuss the possibilities to establish novel metabolic pathways in a microbial cell and evaluate different methods of DNA assembly to obtain expression cassettes for both single enzymes and entire metabolic pathways
4	Understand	Understand and differentiate between in vitro and in vivo biotransformations, including their implications and applications
5	Analyze	Analyze and evaluate integrated process development approaches for upstream and downstream bioprocesses
6	Apply	Apply computational tools and digitalization techniques in bioreactor and bioprocess design, including computer-aided design software and approaches towards SMART biolabs
7	Analyze	Analyze cutting-edge research articles

Indicative Literature

- Wilson, D.B., Sahm, H., Stahmann, K.-P., and Koffas, M. Industrial Microbiology, Wiley-VCH, 2013
- Li, M. and Borodina, I. (2015) Application of synthetic biology for production of chemicals in yeast *Saccharomyces cerevisiae*. FEMS Yeast Research 15: 1–12

- Liu, Z., Zhang, Y. and Nielsen, J. (2019) Synthetic Biology of Yeast. Biochemistry: 58, 1511–1520
- Liese, A., Seelbach K., and Wandrey, C. Industrial Biotransformations, Wiley-VCH, 2006

Entry Requirements

Prerequisites	None
Co-requisites	None
Additional Remarks	None

Assessment and Completion

Components	Examination Type	Duration/ Length	Weight (%)	Minimum	ILOs
Microbial Engineering	Written Examination	60 minutes	100	45%	1-7
Advanced Bioprocess Engineering and Digitalization					

Module Achievements: None

4.2 Scientific Presentations

Module Name	Scientific Presentations
Module Code	2025-MQLS-TS-01
Module ECTS	5
Study Semester	Mandatory status for: - 2025-QLS-MSc 1 Mandatory Elective status for: None
Duration	1 Semester
Program Affiliation	2025-QLS-MSc (Quantitative Life Science)
Module Coordinator(s)	Prof. Dr. Thomas Nugent

Forms of Learning and Teaching		
	Lecture	17.5
	Seminar	17.5
	Tutorial	14
	Independent Study	76
Workload Hours	125 hours	

Module Components	Number	Type	CP
Scientific Presentations	MQLS-TS-01	Lecture and Seminar	5

Module Description

This module consists of two parts: In the first half of the module, all participating faculty will introduce their research with presentations and subsequent discussions. In the second half, students will be asked to present a research publication of their own choice. The first part will enable students to observe different presentation styles and identify their fields of interest in Quantitative Life Science and potential supervisors for the lab rotations. In the second part, students will select a publication in the field of Quantitative Life Science, analyze it, and present it to their classmates. This presentation will focus on the motivation for the study, the different methodologies used, the obtained results, and the performed analyses. The students will also critically discuss the conclusions drawn by the authors.

To prepare students for their presentation, a few lectures will address how to give good scientific presentations, ranging from formal criteria, like the number and design of slides, the use of steady and clear language, and the importance of timing, to content-related topics, e.g., the role of published literature; framing, explaining, and wrapping up an argument; providing conclusions and an outlook of future work. Furthermore, tips pertaining

to audience engagement (eye contact, body language, staying still, etc.), proper handling of (challenging) audience question, etc. will be discussed. During the remainder of the course, each student will give a 30-min presentation on a selected research publication. Each presentation will end with a question-and-answer section, which is followed by instructor feedback on the positive aspects as well as those for further improvement.

To learn how to give a presentation also means one can evaluate a presentation. This will be achieved by each student audience member via the completion of an evaluation form. The instructor will provide corrective feedback if your evaluation is lacking.

Recommended Knowledge

To come with an open mind and flexibly adapt to capture your audience's attention.

Usability and Relationship to other Modules

Master's level study requires students to focus on a particular research area and in doing so you will engage in research. Presentations allow students to bring everything from the classroom to the research laboratory together, and thus prepare for the lab rotations in this study program but also for future job interviews and how to communicate science to other scientists.

Intended Learning Outcomes

No	Competence	ILO
1	Analyse	Analyse and critique scientific publications
2	Organize	Organize ideas and convey them in a convincing, concise, and simple manner during a presentation
3	Demonstrate	Demonstrate how to plan and manage the time allocated to a presentation
4	Prepare	Prepare appealing yet informative slides for the presentation
5	Use	Use proper scientific wording and voice inflection to emphasize important concepts and keep the audience engaged
6	Analyze	Analyze and critique the presentations of their peers

Entry Requirements

Prerequisites	None
Co-requisites	None
Additional Remarks	None

Assessment and Completion

Components	Examination Type	Duration/ Length	Weight (%)	Minimum	ILOs
Scientific Presentations	Presentation		100	45%	All

Module Achievements: Complete evaluations for at least 80% of the peer presentations have to be submitted and fulfil quality criteria defined by the Instructor at the beginning of the course

4.3 Guided Self-study

Module Name	Guided Self-study
Module Code	2025-MQLS-CO-01
Module ECTS	5
Study Semester	Mandatory status for: - 2025-QLS-MSc 1 Mandatory Elective status for: None
Duration	1 Semester
Program Affiliation	2025-QLS-MSc (Quantitative Life Science)
Module Coordinator(s)	Prof. Dr. Felix Jonas

Forms of Learning and Teaching	
Tutorial	35
Exam Preparation	20
Independent Study	70
Workload Hours	125 hours

Module Components	Number	Type	CP
Guided Self Study	MQLS-CO-01	Tutorial	5

Module Description

Quantitative Life Science stands at the intersection of Life Science and Data Science. Successful research in this field necessitates a solid foundation in the life sciences as well as Mathematics and Physics. Hence, basic knowledge in any of these areas is a pre-requisite for most other courses in this program.

In this “Guided Self-study” module, the students will thus, depending on their academic background, recapitulate or acquire the necessary knowledge. The content is divided into three broad areas, Biology, Chemistry and Mathematics/Physics with their own learning goals. These goals are based on the expected knowledge after an undergraduate degree in chemistry, biochemistry or molecular/ cellular biology

Based on a self-test in all three content areas, each student together with a faculty member will build a personalized learning plan. Since most successful candidates will come with expertise in at least one field, usually only one or two areas of focus will require revision or additional learning. All students are expected to take two areas. The student will then study these selected topics in an “inverted classroom” setting. For this, reading materials and other learning materials (videos, exercises) will be provided by the program faculty. In addition, regular tutorial sessions will be offered to enable students to discuss the topics with the

respective subject matter experts. The formation of study groups will be encouraged. In the written examination, only the selected focus areas will be tested for.

Recommended Knowledge

Ability to obtain knowledge independently

Usability and Relationship to other Modules

This module provides the basic knowledge necessary to succeed in most modules of the program

Intended Learning Outcomes

No	Competence	ILO
	Chemistry:	
		<i>Organic Chemistry:</i>
1	Define	Define the structure and properties of carbon-containing compounds
2	Explain	Explain common reaction mechanisms and catalysts
3	Apply	Apply the basics of reaction kinetics to organic reactions
		<i>Physical Chemistry:</i>
4	Discuss	Discuss energetics of chemical reactions
5	Discuss	Discuss chemical reaction rates, diffusion, partition and transport processes
		<i>Analytical Chemistry:</i>
6	Discuss	Discuss quantitative methods and methods for separating and purifying organic/biological compounds
7	Describe	Describe Spectroscopic methods for identifying and characterizing compounds
	Biology:	
		<i>Biochemistry:</i>
1	Explain	Explain the structure, properties and function of biological macromolecules
2	Demonstrate	Demonstrate the principles of enzymes and enzyme kinetics on unknown chemical reactions
3	Remember	Remember basic metabolic pathways (Glycolysis, TCA cycle)
4	Understand	Understand the mechanistic basis of transcription and translation in eukaryotes and prokaryotes
		<i>Cell Biology:</i>
5	Describe	Describe the cellular organization in eukaryotes and prokaryotes
6	Understand	Understand the basics of protein production, targeting, transport and degradation
7	Apply	Apply the principles of regulation of biological process and biological signalling pathways
		<i>Biotechnology:</i>
8	Discuss	Discuss molecular biology techniques and basics of genetic engineering

9	Apply	Apply the fundamentals of material and energy balances to novel bioprocesses
10	Recognize	Recognize the basics of Metabolic engineering
11	Memorize	Memorize bioreactor types and modes
Math/Physics:		
		<i>Math:</i>
1	Understand	Understand the basics of linear algebra and solve simple problems
2	Recognize	Recognize ordinary differential equations and implement numerical solving methods
		<i>Statistics:</i>
3	Interpret	Interpret data using simple descriptive statistics
4	Perform	Perform statistical tests for effect size and significance
5	Explain	Explain basic concepts of Bayesian statistics
		<i>Physics:</i>
6	Explain	Explain and implement the fundamental rules classical mechanics and thermodynamics

Indicative Literature

- Cell Biology: Alberts et al., Molecular Biology of the Cell, Garland Science, latest edition
- Biochemistry: Nelson and Cox, Lehninger Principles of Biochemistry, Freeman Macmillan, latest edition. Stryer et. al., Biochemistry, Freeman Macmillan, latest edition.
- Chemistry: J. Clayden, N. Greeves, S. Warren. Organic Chemistry, 2nd Edition, Oxford University Press, 2012. Zumdahl and Zumdahl, Chemistry, Brooks Cole, latest edition;
- Math / Statistics: E. N. Bodine, S. Lenhart, and L. J. Gross. Mathematics for the Life Sciences. Princeton University Press, latest edition
- Physics: Zinke-Allmang et al., Physics for the Life Sciences, Nelson Education, latest edition.

Entry Requirements

Prerequisites	None
Co-requisites	None
Additional Remarks	None

Assessment and Completion

Components	Examination Type	Duration/ Length	Weight (%)	Minimum	ILOs
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Guided Self Study	Written Examination	120 minutes (2 x 60 minutes)	100	45%	All IOLs of selected areas
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Module Achievements: None

4.4 Data Tools for the Life Sciences

Module Name	Data Tools for the Life Sciences
Module Code	2025-MQLS-CO-02
Module ECTS	5
Study Semester	Mandatory status for: - 2025-QLS-MSc 1 Mandatory Elective status for: None
Duration	1 Semester
Program Affiliation	2025-QLS-MSc (Quantitative Life Science)
Module Coordinator(s)	Prof. Dr. Felix Jonas

Forms of Learning and Teaching	
Lecture	17.5
Tutorial	17.5
Exam Preparation	10
Independent Study	80
Workload Hours	125 hours

Module Components	Number	Type	CP
Data Tools for the Life Sciences - Lecture	MQLS-CO-02-A	Lecture/Tutorial	2.5
Data Tools for the Life Sciences - Project	MQLS-CO-02-B	Project	2.5

Module Description

The recent developments in the field of experimental life science allow researchers to collect data at an unprecedented scale and complexity not directly comprehensible to even educated scientists. Generating, analyzing, and concluding from these large data sets is at the heart of Quantitative Life Science and usually relies on modern computational tools.

Thus, this module provides the student with the fundamental skills for computer assisted processing, analyzing and visualizing large data sets from fields of biology and chemistry. A particular focus will be on multi-dimensional, numerical data common to most experiment types and usually stored in matrix format. Other more specific data types, like DNA sequences, protein sequences, and protein structures, will also be discussed. Moreover, in addition to classical data analysis, the students will also learn how to leverage large-language models to facilitate their interaction with computers.

The course will consist of lectures to introduce the students to the principles, guided tutorials, where the student can apply their skills to given problems, and homework assignment where the students apply their skills to selected case studies in a collaborative or individual settings. In terms of computational data analysis, the students will learn how to construct hypothesis, and to select and apply advanced statistical tests to verify them. They will also learn how to utilize descriptive statistics to provide an informative overview of data. Additionally, the students will also acquire the basics of data fitting, dimensionality reduction, and machine learning to identify patterns in the data.

With respect to visualization, the student will be introduced to the diverse set of plot types, such as bar graphs, scatter plots, and heatmaps, and learn when and how to apply them. A particular focus will be on visualizing multidimensional data and using different scales and normalization techniques to match the nature of the underlying data and the desired hypothesis.

The topic of data processing will introduce the students to the concept of automated data processing routines, also known as “pipelines”. as well as equip them with the skills to build simple pipelines and understand more complex ones to convert experimental raw data sets from chemistry and biology into numerical matrixes. Furthermore, the students will be exposed to ethical aspects and best practices of data analysis.

Recommended Knowledge

Install coding program (e.g. Python) and familiarize yourself with the basic data types and syntax.

Usability and Relationship to other Modules

The methods and concepts learned in this module can be used in research projects as well as the master thesis. Moreover, the computational skills can be applied in several core and elective modules (e.g. Omics I and II, Quantitative Cell Biology etc.).

Intended Learning Outcomes

No	Competence	ILO
1	Discuss	Discuss the fundamental principles of machine learning
2	Implement	Implement dimensionality reduction and data fitting for large, multidimensional datasets
3	Compare	Compare the advantages and limits of different statistical tests and graph types
4	Examine	Examine and question common data processing routines in the Life Science
5	Select	Select and defend different descriptive statistics, visualization techniques and statistical tests to highlight or examine certain data aspects

6	Critique	Critique the use of different analysis and visualization techniques in the published papers
7	Collaborate	Collaborate effectively, assume responsibility and leadership

Indicative Literature

- McKinney Wes, Python for Data Analysis: Data Wrangling with pandas, NumPy, and Jupyter, 3rd Edition, New York: Oreilly Media, 2022
- David Landup, Data Visualization in Python with Pandas and Matplotlib, Stack Abuse, 2021
- Python Data Science Handbook: Essential Tools for Working with Data, 2nd Edition, O'Reilly Media Inc., 2022
- Statistics and Data Visualization with Python (Chapman & Hall/CRC The Python Series), 1st Edition, 2023

Entry Requirements

Prerequisites	None
Co-requisites	None
Additional Remarks	None

Assessment and Completion

Components	Examination Type	Duration/ Length	Weight (%)	Minimum	ILOs
Data Tools for the Life Sciences - Lecture	Written Examination	60 minutes	50	45%	1, 3, 4, 6
Data Tools for the Life Sciences - Project	Project Report	Approx. 8 pages	50	45%	2, 3, 5, 7

Module Achievements: None

4.5 Laboratory Rotation QLS I

Module Name	Laboratory Rotation QLS I
Module Code	2025-MQLS-RES-01
Module ECTS	10
Study Semester	Mandatory status for: - 2025-QLS-MSc 1 Mandatory Elective status for: None
Duration	1 Semester
Program Affiliation	2025-QLS-MSc (Quantitative Life Science)
Module Coordinator(s)	SPC

Forms of Learning and Teaching	
Seminar	17.5
Laboratory	182.5
Exam Preparation	20
Independent Study	30
Workload Hours	250 hours

Module Components	Number	Type	CP
Laboratory Rotation QLS I -lab	MQLS-RES-01-A	Laboratory	7.5
Laboratory Rotation QLS I -Seminar	MQLS-RES-01-B	Seminar	2.5

Module Description

The laboratory rotations are part of the research focus of the Quantitative Life Science program. Students select a research field and supervisor among all participating faculty based on their knowledge and interest. In their chosen field (e.g. Medicinal chemistry, analytical chemistry, biochemistry, cell biology, computational chemistry etc.) students will receive intense training on experimental procedures, software, coding, instrument operation from the members of the research group selected. Following training they will receive a small research project to work. Throughout the project, discuss their results with their supervisor.

Recommended Knowledge

- Attendance of laboratory safety instructions. Background literature for preparation provided by

individual faculty.

-Basic laboratory skills

Attendance of safety instructions.

Usability and Relationship to other Modules

The module prepares for the following two laboratory rotations II & III and the Master thesis.

Additionally, the analyses carried out might relate to other Core and elective modules e.g. work in analytical chemistry to Metabolomics and Structure elucidation or work in bioinformatics to Machine Learning or Advanced Molecular Simulations.

Intended Learning Outcomes

No	Competence	ILO
1	Plan	Plan and conduct wet- or dry-lab experiments in a Life Science research area where background knowledge exists
2	Demonstrate	Demonstrate ability to operate scientific instrumentation and master essential software
3	Produce	Produce experimental data or perform computational dataset research
4	Interpret	Interpret and analyze experimental data
5	Discuss	Discuss and present their data and evaluate their significance

Indicative Literature

- Provided by individual faculty depending on their field of research

Entry Requirements

Prerequisites	None
Co-requisites	Scientific Presentations
Additional Remarks	None

Assessment and Completion

Components	Examination Type	Duration/ Length	Weight (%)	Minimum	ILOs
Laboratory Rotation QLS I - lab	Laboratory Report	10-15 pages	75	45%	1-4
Laboratory Rotation QLS I - Seminar	Presentation	20 minutes	25	45%	4,5

Module Achievements: None

4.6 Structure Elucidation

Module Name	Structure Elucidation
Module Code	2025-MQLS-CHE-02
Module ECTS	5
Study Semester	Mandatory status for: None Mandatory Elective status for: - 2025-QLS-MSc 2 - 2025-AdMat-MSc 2
Duration	1 Semester
Program Affiliation	2025-QLS-MSc (Quantitative Life Science)
Module Coordinator(s)	Prof. Dr. Nikolai Kuhnert Prof. Dr. Ulrich Kortz

Forms of Learning and Teaching	
Lecture	35
Exam Preparation	20
Independent Study	70
Workload Hours	125 hours

Module Components	Number	Type	CP
Structure Elucidation	MQLS-CHE-02	Lecture	5

Module Description

Modern Life Science and Material Science operate at the molecular level. The individual chemical structure defines the properties of biomolecules and materials and interactions. Hence it forms the basis of life and of material science. A thorough understanding of instrumental methods used for chemical structure elucidation in solution, in the solid state and in the gas phase, and their application to “known” and novel “unknown” substances is therefore an essential part of both MSc programs.

The module builds on previous knowledge at the Bachelor level and gives an intermediate level introduction and actual exposure to key experimental techniques such as nuclear magnetic resonance (NMR) spectroscopy, single crystal X-ray diffraction, infrared (IR) spectroscopy, mass spectrometry, Mössbauer spectroscopy and electron paramagnetic resonance (EPR) spectroscopy. Typically, a chemical structure elucidation process requires the use of a combination of several experimental techniques, which will be presented to the students along with applied exercises. The module will further introduce more advanced techniques such as multidimensional NMR spectroscopy (homonuclear and heteronuclear) of organic and

inorganic compounds, materials, metabolites and proteins, high resolution tandem mass spectrometry, and single crystal as well as powder X-ray diffraction combined with data treatment and molecular visualization using state of

the art software.

Recommended Knowledge

- Background literature for preparation provided by individual faculty.

- Basic knowledge on chemical bonding and

spectroscopic techniques (BSc level

Analytical Methods and Inorganic Chemistry)

Understanding of chemical structure and

bonding

Intended Learning Outcomes

No	Competence	ILO
1	Appreciate	Appreciate information content of spectroscopic and diffraction data
2	Predict	Predict and propose chemical structures from spectroscopic and diffraction data
3	Predict	Predict and deduce spectral data from chemical structures
4	Interpret	Interpret spectral data and assign signals to molecular features
5	Recommend	Recommend and rate spectroscopic experiments with a view of solving ambiguous chemical structure problems

Indicative Literature

- Sternhill and Kalman 2D NMR spectroscopy
- Sternhill and Kalmann Solving organic spectroscopic problems
- David W. H. Rankin, Norbert W. Mitzel, Carole A. Morrison: Structural Methods In Molecular Inorganic Chemistry, 2nd edition, Wiley, 2013

Entry Requirements

Prerequisites	Guided Self-study
Co-requisites	Experimental Techniques
Additional Remarks	

Assessment and Completion

Components	Examination Type	Duration/ Length	Weight (%)	Minimum	ILOs
Structure Elucidation	Written Examination	120 minutes	100	45%	All

Module Achievements: None

4.7 Experimental Techniques

Module Name	Experimental Techniques
Module Code	2025-MQLS-CO-03
Module ECTS	5
Study Semester	Mandatory status for: - 2025-QLS-MSc 2 Mandatory Elective status for: None
Duration	1 Semester
Program Affiliation	2025-QLS-MSc (Quantitative Life Science)
Module Coordinator(s)	Prof. Dr. Torsten John

Forms of Learning and Teaching	
Lecture	35
Exam Preparation	20
Independent Study	70
Workload Hours	125 hours

Module Components	Number	Type	CP
Experimental Techniques	MQLS-CO-03	Lecture	5

Module Description

Modern experimental Life Science heavily depends on data generation using a combination of very diverse techniques. The module aims to provide the student with a toolbox of key experimental methods that can address a multitude of diverse biological questions. Therefore, the course will discuss the theoretical background, the experimental protocol, the analysis framework as well as the strengths and weaknesses of each technique.

The methods and techniques presented in this module were selected according to the following

criteria: 1. Relevance of key techniques that drove progress in Life Sciences over the last decades;

2. Techniques available at Constructor University Laboratories and available for Research

Laboratory Components; 3. No overlap with techniques discussed in depth in other modules (e.g. mass spectrometry in Omics II or sequencing in Omics I).

Notably, key advances in published research will lead to constant revision and update of the

techniques presented in this module since modern sciences' key driver is experimental technology.

Techniques presented include the following:

- Methods for studying energetics of biochemical processes (Microcalorimetry, isothermal titration calorimetry ITC)
- Fluorescence-based methods (FRET, FACS, single photon etc.) including labelling strategies for incorporating fluorescent labels to biomolecules
- Spectroscopic techniques including NMR-, CD-, IR-, Raman-spectroscopy and their adaptation in microscopy.
- Separation techniques including separation of biomolecules (chromatography, electrophoresis, centrifugation etc.) and cell separation (flow cytometry)
- Methods for studying bio-molecular interactions, e.g. atomic force microscopy (AFM), surface plasmon resonance (SPR), quartz crystal microbalances (QCM), nano differential fluorimetry (nano-DSF)
- Cell culture methods, e.g. Primary co-culture or induced pluripotent stem cells.

Recommended Knowledge

- Background literature for preparation provided by individual faculty.
- Basic knowledge on separation, identification and quantification of biomolecules. Ability to generate and interpret experimental data

Usability and Relationship to other Modules

Content, techniques and methods learned in all Quantitative Life Science core, elective and transferrable skill modules will be applied to solve a real significant research question. Additional key experimental techniques form focus of other modules e.g. mass spectrometry (Omics II), sequencing (Omics I), microscopy (quantitative cell biology) or NMR spectroscopy and XRD diffraction (structure elucidation).

Intended Learning Outcomes

No	Competence	ILO
1	Examine	Examine information content of experimental data

2	Apply	Apply appropriate experimental techniques for solving a scientific problem
3	Appraise	Appraise and compare the scope and limitations of different experimental methods and techniques
4	Explain	Explain the theoretical and practical aspects of the key experimental methods in Life Sciences

Indicative Literature

- Lottspeich and Engels, Bioanalytics, Wiley-VCH.

Entry Requirements

Prerequisites	Guided Self-study
Co-requisites	None
Additional Remarks	None

Assessment and Completion

Components	Examination Type	Duration/ Length	Weight (%)	Minimum	ILOs
Experimental Techniques	Written Examination	120 minutes	100	45%	All

Module Achievements: None

4.8 Omics I - Genomics, Transcriptomics

Module Name	Omics I - Genomics, Transcriptomics
Module Code	2025-MQLS-CO-04
Module ECTS	5
Study Semester	Mandatory status for: - 2025-QLS-MSc 2 Mandatory Elective status for: None
Duration	1 Semester
Program Affiliation	2025-QLS-MSc (Quantitative Life Science)
Module Coordinator(s)	Prof. Dr. Felix Jonas Prof. Dr. Marc-Thorsten Hütt

Forms of Learning and Teaching	
Lecture	35
Exam Preparation	20
Independent Study	70
Workload Hours	125 hours

Module Components	Number	Type	CP
Omics I - Genomics, Transcriptomics Lecture	MQLS-CO-04	Lecture	5

Module Description

Since their identification as the main information carrier in any biological system, nucleic acids (DNA and RNA) have attracted researchers' interest for diverse purposes. While initially prohibitively expensive and time-consuming, the advent of next generation sequencing reduced the cost and time for DNA sequencing. Now, whole genomes and transcriptomes of organisms are accessible to research but demand new quantitative approaches for their analysis.

In this course, students will learn how to obtain and analyze data from three main omics categories, genomics, epigenomics, and transcriptomics, to answer fundamental and applied questions in the field of life science. Based on examples from recent scientific literature, the students will delve into the methods and data types associated with this field and discuss their interpretation, and their biological implications. Moreover, the homework assignment will allow the students to apply these new skills.

An omics project often follows a common strategy: Starting with experimental design, and data gathering, the data are then analyzed using established bioinformatics methods and integrated with existing information in databases to draw functional conclusions. An example is the defining the phenotypic state of cells in a disease from transcriptomics data: First RNA sequencing is performed to determine the transcriptome in healthy and affected individuals, then the sequencing reads are mapped to specific genes, which are subsequently compared between both groups (cases and healthy controls) in the cohort. Statically significant genes are then defined using established methods (like DESeq2) and, for example, integrated with information in the Gene Ontology (GO) database. The GO database contains assignments of categories of molecular function to each gene and can, with a few caveats, be used as a steppingstone towards a functional interpretation of the found gene sets.

Such paths from experiment and data to functional conclusions, as well as the most prominent examples of each step, will be highlighted in this course. We will provide insight into the established and advanced techniques in genomics, epigenomics and transcriptomics, e.g. Whole-Genome- Sequencing, ChIP-seq and single-cell RNAseq, as well as common bioinformatics tools used in this context and the theory behind them. We will also discuss relevant databases, the data formats they employ and the computational tools available for using them.

This collection of case studies and examples will provide a solid foundation for a deep practical understanding of this emerging field; which biological questions can be addressed with which combination of omics technology, databases, and computational methods.

Recommended Knowledge

- Recapitulate the concepts and mechanisms of molecular genetics.

- Basic knowledge of Molecular Biology,

Biochemistry and Bioinformatics.

Usability and Relationship to other Modules

This module is conceptionally linked to the Omics II Module, as it focuses on DNA and RNA, while the latter one focuses on Proteins and Metabolites. The experiments and analysis techniques introduced in this module will also support the topics discussed in the elective modules or find application in the research rotations or the master thesis.

Intended Learning Outcomes

No	Competence	ILO
1	Recall	Recall key concepts in genomics, epigenomics, and transcriptomics
2	Explain	Explain the steps involved in an omics experiment from experimental design to functional conclusions
3	Apply	Apply bioinformatics methods and integrated existing databases to analyze omics datasets and interpret the results

4	Compare	Compare and contrast different bioinformatics tools, omics technologies and databases and their suitability for various omics analyses
5	Design	Design an experimental workflow for a specific biological question using omics techniques, relevant databases, and computational tools
6	Assess	Assess the validity and reliability of omics data, analysis and biological interpretation
7	Formulate	Formulate recommendations for best practices in omics data analysis based on case studies and recent literature
8	Collaborate	Collaborate effectively in group projects to analyze, interpret, and present omics data

Entry Requirements

Prerequisites	Guided Self-study Data Tools for the Life Sciences
Co-requisites	None
Additional Remarks	

Assessment and Completion

Components	Examination Type	Duration/ Length	Weight (%)	Minimum	ILOs
Omics I - Genomics, Transcriptomics Lecture	Written Examination	120 minutes	100	45%	All

Module Achievements: Homework assignments have to be passed by an average of 45%.

4.9 Scientific Writing

Module Name	Scientific Writing
Module Code	2025-MQLS-TS-02
Module ECTS	5
Study Semester	Mandatory status for: - 2025-QLS-MSc 2 - 2025-AdMat-MSc 2 Mandatory Elective status for: None
Duration	1 Semester
Program Affiliation	2025-QLS-MSc (Quantitative Life Science)
Module Coordinator(s)	Prof. Dr. Katrin Rosenthal

Forms of Learning and Teaching	
Seminar	35
Exam Preparation	35
Independent Study	55
Workload Hours	125 hours

Module Components	Number	Type	CP
Scientific Writing	MQLS-TS-02	Seminar	5

Module Description

This module is designed to provide students with the skills and knowledge required for effective scientific writing. The module will cover the principles of clear and concise writing, structuring scientific papers and research proposals, presenting data and results, and understanding the submission and publication process. By the end of this module, students will be able to create well-organized, coherent, and persuasive scientific documents. The seminar will enhance critical thinking skills by evaluating scientific arguments and methodologies. The ability to gather relevant information and scientific references will be improved. The seminars provide theoretical foundations and examples of scientific writing with practical sessions for writing and revising texts.

Intended Learning Outcomes

No	Competence	ILO
1	Understand	Understand key components of scientific papers, structure and standards in scientific writing, including abstracts, introductions, methods, results, and discussions and compare different types of scientific documents, such as research articles, reviews, and case studies

2	Apply	Apply principles of effective scientific writing to produce clear and concise sentences and paragraphs and use of appropriate formatting and citation styles in scientific documents
3	Understand	Understand the purpose and importance of scientific writing and scientific communication and understand the publication process and peer review process
4	Analyze	Analyze and evaluate data presentation, tables and figures, as well as data interpretation to identify strengths and weaknesses in writing style and structure and evaluate scientific papers for logical flow, clarity, and adherence to conventions
5	Apply	Apply their theoretical knowledge on a writing a short proposal about their own research
Additional Learning Outcomes (Interdisciplinary Skills)		
6	Develop	Develop strong written communication skills that are applicable in both academic and professional settings
7	Understand	Understand and apply principles of ethical writing, including avoiding plagiarism and ensuring proper attribution
8	Promote	Promote a sense of integrity and responsibility in reporting and discussing scientific findings

Indicative Literature

- Glasman-Deal H. (2009). Science Research Writing. Imperial College Press
- Morris J.R., Jehn T.R., Vaughan C., Pantages E., Torello T., Bucheli M., Lohman D., Lue R., Harvard University (2007). A Student's Guide to Writing in the Life Sciences. Harvard University

Entry Requirements

Prerequisites	None
Co-requisites	None
Additional Remarks	None

Assessment and Completion

Components	Examination Type	Duration/ Length	Weight (%)	Minimum	ILOs
Scientific Writing	Term Paper	5 pages	100	45%	All

Module Achievements: None

4.10 Laboratory Rotation QLS II

Module Name	Laboratory Rotation QLS II
Module Code	2025-MQLS-RES-02
Module ECTS	10
Study Semester	Mandatory status for: - 2025-QLS-MSc 2 Mandatory Elective status for: None
Duration	1 Semester
Program Affiliation	2025-QLS-MSc (Quantitative Life Science)
Module Coordinator(s)	SPC

Forms of Learning and Teaching	
Laboratory	17.5
Laboratory	182.5
Exam Preparation	20
Independent Study	30
Workload Hours	250 hours

Module Components	Number	Type	CP
Lab rotation QLS II - Lab	MQLS-RES-02-A	Seminar	7.5
Lab rotation QLS II - Seminar	MQLS-RES-02-B	Seminar	2.5

Module Description

The laboratory rotations are part of the research focus of the Quantitative Life Science program. Students select a research field and supervisor among all participating faculty of their choice and interest. In their chosen field (e.g. Medicinal chemistry, analytical chemistry, biochemistry, cell biology, computational chemistry etc.) students will receive intensive training on experimental procedures, software, coding, instrument operation from the members of the research group selected. Following training they will receive a small research project to work on. The second laboratory rotation should be completed in a different research group with a different supervisor than the previous laboratory rotation I. This will expose the student to an alternative area of science and research style. Furthermore, it should allow for the (optional) development of collaborative research projects between different research groups in the Master thesis.

Recommended Knowledge

- Attendance of laboratory safety instructions. Background literature for preparation provided by

individual faculty.

- Basic laboratory skills

Attendance of safety instructions

Usability and Relationship to other Modules

In combination with rotation I, the module prepares students for the laboratory rotations III and the Master thesis. Additionally, the research projects may relate to other Core and elective modules e.g. work in analytical chemistry to Metabolomics and Structure elucidation or work in bioinformatics to Machine Learning or Advanced Molecular Simulations.

Intended Learning Outcomes

No	Competence	ILO
1	Independently	Independently acquire necessary background knowledge in a new area of research
2	Plan	Plan and conduct experiments in a new area of research (i.e., other than rotation I)
3	Produce	Produce data sets in a Life Science research area
4	Interpret	Interpret and analyze experimental and computational data
5	Discuss	Discuss and present their experimental data and evaluate their significance

Indicative Literature

- Provided by individual faculty depending on their field of research

Entry Requirements

Prerequisites	None
Co-requisites	None
Additional Remarks	None

Assessment and Completion

Components	Examination Type	Duration/ Length	Weight (%)	Minimum	ILOs
Lab rotation QLS II - Lab	Laboratory Report	10-15 pages	75	45%	1-4
Lab rotation QLS II - Seminar	Presentation	20 minutes	25	45%	4,5

Module Achievements: None

4.11 Quantitative Cell Biology

Module Name	Quantitative Cell Biology
Module Code	2025-MQLS-CEL-01
Module ECTS	5
Study Semester	Mandatory status for: None Mandatory Elective status for: - 2025-QLS-MSc 3
Duration	1 Semester
Program Affiliation	2025-QLS-MSc (Quantitative Life Science)
Module Coordinator(s)	Prof. Dr. Klaudia Brix

Forms of Learning and Teaching	
Seminar	17.5
Tutorial	17.5
Exam Preparation	20
Independent Study	70
Workload Hours	125 hours

Module Components	Number	Type	CP
Quantitative Cell Biology seminar	MQLS-CEL-01-A	Seminar	2.5
Quantitative Cell Biology tutorial	MQLS-CEL-01-B	Tutorial	2.5

Module Description

Complex organisms, like humans, consists of a multitude of different cell types with countless interactions, which are fundamental to all physiologic and pathologic processes. The module “Quantitative Cell Biology” aims at teaching students tools to resolve this heterogeneity of cell and tissue composition at the molecular level. For example, in microscope experiments well-defined biomarkers of specific intra- and extracellular components are stained and cells and tissues are imaged at high resolution. Multiplexing (using a broad range of markers) allows assessing cell and tissue heterogeneity in an integrative fashion. Cell types and tissue structures as well as biomarkers thereof will be introduced in the seminar. This knowledge will be applied during the practical tutorials, for example by performing microscopical inspection of stained specimen. Furthermore, the students will learn how to use computational tools to objectively, reproducibly and rapidly examine and analyse microscope images in the age of high-content microscopy. Thus, the goal is to enable students to analyze cells and tissues qualitatively and quantitatively, thereby gaining information on their respective molecular

composition and state. The participants will present results of individually chosen experimental settings as a module achievement.

Recommended Knowledge

- Make yourself familiar with the different cell types and tissues of the human body
- Read about common and rare diseases mentioned in the cell biological textbook and in recent literature (follow recommendations given during seminars; search: <https://pubmed.ncbi.nlm.nih.gov/>; <https://medlineplus.gov/>) focus on cancer or endocrinological disorders
- Familiarize yourself with image analysis tools (e.g. <https://cellprofiler.org/>; <https://tutorials.cellprofiler.org/>)
- Familiarize yourself with experimental “cell-painting” tools and molecular probes used as markers of cellular structures and compartments (ThermoFisher-Invitrogen; Broad Institute)
- Visit journal clubs or lab meetings of the research groups with partial cell biological orientation
- Visit the Molecular Life Sciences seminar series where researchers from other institutions present
- Basic knowledge of cell biology methods and concepts; ability to read and understand scientific articles; critical discussion skills

Intended Learning Outcomes

No	Competence	ILO
1	Understand	Understand molecular composition of cells and tissues
2	Apply	Apply the concept of biomarkers
3	Analyse	Analyse quantitative and qualitative changes of biomolecules in disease
4	Apply	Apply image analysis tools
5	Critically	Critically interpret quantitative changes in cellular biomolecules
6	Enhance	Enhance personal competence in communicating and validating scientific data in the form of a comprehensive results presentation

Indicative Literature

- Alberts, B., et al., Molecular Biology of the Cell, 7th Edition, W.W. Norton & Company, Inc., 2022
- Pollard, T.D., Earnshaw, W.C., et al., Cell Biology, 4th Edition, Elsevier, 2022

- Milo, R., Phillips, R., Cell Biology by the Numbers, Garland Science, Taylor & Francis Group, LLC, 2016
- CellProfiler™, Free open-source software for measuring and analyzing cell images. Download from cellprofiler.org

Entry Requirements

Prerequisites	None
Co-requisites	Guided Self-study Data Tools for the Life Sciences
Additional Remarks	

Assessment and Completion

Components	Examination Type	Duration/ Length	Weight (%)	Minimum	ILOs
Quantitative Cell Biology seminar	Written Examination	60 minutes	50	45%	1,2,3
Quantitative Cell Biology tutorial	Presentation	20 minutes	50	45%	4,5,6

Module Achievements: None

4.12 Laboratory Rotation QLS III

Module Name	Laboratory Rotation QLS III
Module Code	2025-MQLS-RES-03
Module ECTS	10
Study Semester	Mandatory status for: - 2025-QLS-MSc 3 Mandatory Elective status for: None
Duration	1 Semester
Program Affiliation	2025-QLS-MSc (Quantitative Life Science)
Module Coordinator(s)	SPC

Forms of Learning and Teaching	
Seminar	17.5
Laboratory	182.5
Exam Preparation	20
Independent Study	30
Workload Hours	250 hours

Module Components	Number	Type	CP
Lab Rotation QLS III - Lab	MQLS-RES-03-A	Laboratory	7.5
Lab Rotation QLS III – Seminar	MQLS-RES-03-B	Seminar	2.5

Module Description

The laboratory rotations are part of the research focus of the Quantitative Life Science program. It is recommended but not required that the Laboratory rotation III will take place in the laboratory of a faculty member already selected in Laboratory rotation I or II. Ideally, the Master thesis module will be carried out in the laboratory selected for Laboratory rotation III to allow for a more comprehensive analysis of the research topic. In this context, the Lab Rotation III by researching a more advanced topic with a higher degree of independence directly prepares for subsequent Master thesis. A laboratory rotation in an external laboratory is possible on special application, if supervised by a School of Science faculty member at Constructor University. Approval must be given by the SPC.

Recommended Knowledge

- Attendance of laboratory safety instructions. Background literature for preparation provided by individual faculty.

- Basic laboratory skills

Attendance of safety instructions

Usability and Relationship to other Modules

The module prepares for the Master thesis as the participants are expected to show a higher level of independence in research strategy development. Additionally, the skills acquired and deepened in rotation III will help in designing and conducting the independent research project for the master thesis.

Intended Learning Outcomes

No	Competence	ILO
1	Independently	Independently plan and conduct experiments with a larger strategic perspective
2	Independently	Independently establish new experimental protocols or analysis algorithms
3	Produce	Produce experimental data / computational datasets in the Life Science
4	Solve	Solve experimental problems based on data obtained
5	Read	Read and implement information from scientific literature
6	Interpret	Interpret and analyze experimental and computational data in the Life Sciences
7	Discuss	Discuss and present their experimental data and evaluate their significance

Indicative Literature

- Provided by individual faculty depending on their field of research

Entry Requirements

Prerequisites	None
Co-requisites	None
Additional Remarks	None

Assessment and Completion

Components	Examination Type	Duration/ Length	Weight (%)	Minimum	ILOs
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Lab Rotation QLS III - Lab	Laboratory Report	10-15 pages	75	45%	1-4
Lab Rotation QLS III – Seminar	Presentation	20 minutes	25	45%	4,5

Module Achievements: None

4.13 Computational Life Science - Modelling and Simulations

Module Name	Computational Life Science - Modelling and Simulations
Module Code	2025-MQLS-CO-06
Module ECTS	5
Study Semester	Mandatory status for: - 2025-QLS-MSc 3 Mandatory Elective status for: None
Duration	1 Semester
Program Affiliation	2025-QLS-MSc (Quantitative Life Science)
Module Coordinator(s)	Prof. Dr. Marc-Thorsten Hütt Prof. Dr. Katrin Rosenthal

Forms of Learning and Teaching	
Lecture	35
Exam Preparation	20
Independent Study	55
Workload Hours	110 hours

Module Components	Number	Type	CP
Computational Life Science - Modelling and Simulations	MQLS-CO-06	Lecture	5

Module Description

The course deals with the understanding of microbial cells and biocatalytic reaction networks as production systems for the synthesis of various products. At the core of these practical applications are fundamental questions involving mathematical modelling, simulation, and the computational analysis of metabolic systems. We will introduce the basic ideas of kinetic modelling and models of enzyme kinetics. We will illustrate how elementary ideas from linear algebra can be employed to simulated metabolic fluxes (a concept called flux-balance analysis) and how some biological conclusions about metabolic systems can already be drawn from the metabolic network alone. In this course we will discuss this approach and highlight the graph-theoretical methods enabling it.

As these descriptions – from the biological network as a black box to flux-balance analysis and mechanistic modelling – all employ very different levels of detail and have distinct computational challenges, it is important to discuss, which description is best suited for a given application and how the different descriptions fit together. A high degree of practical

experience through (dry) laboratory and computer work ensures the ability to apply the acquired theoretical knowledge.

Recommended Knowledge

Fundamentals of Biochemistry and Metabolic

Pathways.

Intended Learning Outcomes

No	Competence	ILO
1	Understand	Understand the basic concepts of findable, accessible, interoperable, and reproducible (FAIR) data and data standardization and evaluate their importance for data exchange between disciplines, as well as experimental and dry-lab research
2	Understand	Understand computational design of bioreactions and apply methods and tools used for the computational planning and optimization of biochemical reactions
3	Understand	Understand the basics of mechanistic (based on the understanding of the underlying biological mechanisms) and empirical modelling (based on observed data) of biochemical reaction networks
4	Apply	Apply and analyze different modelling techniques based on, e.g., ordinary differential equations (ODEs), one-factor-at-a-time (OFAT) experiments, and Bayesian optimization
5	Application	Application of specialized software to simulate and analyze biochemical networks and their dynamics, facilitating the understanding and prediction of their behavior. Evaluate the results and the biological conclusions that can be drawn from them
6	Additional	Additional Learning Outcomes (Interdisciplinary Skills):
7	Collaborate	Collaborate effectively in group projects to model and simulate metabolic systems
8	Communicate	Communicate complex concepts of metabolic modelling and simulation clearly in presentations

Indicative Literature

- Klipp, E., Herwig, R., Kowald, A., Wierling, C., & Lehrach, H. (2005). Systems biology in practice: concepts, implementation and application. John Wiley & Sons.
- Carbonell, P (2019), Metabolic Pathway Design: A Practical Guideline. Springer Nature Switzerland AG

Entry Requirements

Prerequisites	Guided Self-study
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	Data Tools for the Life Sciences
Co-requisites	None
Additional Remarks	

Assessment and Completion

Components	Examination Type	Duration/ Length	Weight (%)	Minimum	ILOs
Computational Life Science - Modelling and Simulations	Written Examination	120 minutes	100	45%	All

Module Achievements: None

4.14 Molecular Simulations

Module Name	Molecular Simulations
Module Code	2025-MQLS-COM-01
Module ECTS	5
Study Semester	Mandatory status for: None Mandatory Elective status for: - 2025-QLS-MSc 3
Duration	1 Semester
Program Affiliation	2025-QLS-MSc (Quantitative Life Science)
Module Coordinator(s)	Prof. Dr. Ulrich Kleinekathöfer

Forms of Learning and Teaching	
Lecture	17.5
Laboratory	17.5
Exam Preparation	20
Independent Study	60
Workload Hours	115 hours

Module Components	Number	Type	CP
Molecular Simulations Lecture	MQLS-COM-01-A	Lecture	2.5
Molecular Simulations Lab	MQLS-COM-01-B	Laboratory	2.5

Module Description

This course will provide the students with an introductory level understanding of the concepts and techniques for the modelling and simulations of biological systems on the molecular scale. Both fundamentals and applications of the most popular molecular simulation methods especially Molecular Dynamics (MD) approaches will be discussed, through examples distilled from the frontier of computational molecular life sciences. Integrated practical work in the computer lab will closely follow the theoretical discussion during the lecture. Computational hands-on problems using state-of-the-art scientific software packages are done partially in supervised sessions and partially as self-study.

Recommended Knowledge

- Students taking this course should have some general knowledge of basics physics and chemistry.
- Basic knowledge of math, physics and chemistry.

Intended Learning Outcomes

No	Competence	ILO
1	Explain	Explain the theory and computational techniques of (bio)molecular dynamics simulations in the realm of (bio)molecular systems
2	Students	Students will be familiar with software for molecular dynamics simulations
3	Apply	Apply modern molecular simulation techniques to practical problems with a focus on Life Sciences
4	Analyze	Analyze, visualize and present results obtained from molecular simulations

Indicative Literature

- Alavi, S., P, Molecular Simulations: Fundamentals and Practice, Wiley-VCH, Weinheim, 2020
- Frenkel, D., and Smit, B., Understanding Molecular Simulation, 3rd Edition, Academic Press, 2023

Entry Requirements

Prerequisites	Guided Self-study Data Tools for the Life Sciences
Co-requisites	None
Additional Remarks	

Assessment and Completion

Components	Examination Type	Duration/ Length	Weight (%)	Minimum	ILOs
Molecular Simulations Lecture	Written Examination	90 minutes	50	45%	1,4
Molecular Simulations Lab	Laboratory Report	5-10 pages	50	45%	2-4

Module Achievements: None

4.15 Omics II - Microbiomics, Proteomics and Metabolomics

Module Name	Omics II - Microbiomics, Proteomics and Metabolomics
Module Code	2025-MQLS-CO-05
Module ECTS	5
Study Semester	Mandatory status for: - 2025-QLS-MSc 3 Mandatory Elective status for: None
Duration	1 Semester
Program Affiliation	2025-QLS-MSc (Quantitative Life Science)
Module Coordinator(s)	Prof. Dr. Matthias Ullrich Prof. Dr. Nikolai Kuhnert

Forms of Learning and Teaching	
Lecture	35
Exam Preparation	20
Independent Study	70
Workload Hours	125 hours

Module Components	Number	Type	CP
Omics II - Microbiomics, Proteomics, and Metabolomics	MQLS-CO-05	Lecture	5

Module Description

Modern Life Science has defined the sciences of Omics as methods for the global analysis of the most important groups of molecules. In Omics II, the investigation of the totality of microbial life (microbiomics), proteins (proteomics) and small molecule metabolites (metabolomics) of any given living organism or ecosystem forms the focus of the module. The module consists of two sections: Microbiomics and Proteomics/Metabolomics.

Microbiomics, proteomics and metabolomics depend on either deep nucleotide sequencing or high-resolution tandem mass spectrometry, respectively, as the key experimental techniques. Hence the two sub-modules will start with an introduction to these technique, experimental methods for genomic DNA extraction, protein isolation and purification, type of data generated and techniques and strategies (targeted versus shotgun) for mass spectrometry data interpretation both manually and by using software databases and algorithms.

A key element of Microbiomics is the definition and identification of the actual microbial ecosystem to be investigated. Likewise, a solid understanding of taxon units as well as differences

between pro- and eukaryotes is needed. Amplicon sequencing as well as metagenome sequencing will be introduced. Ultimately, the course participants need to understand the basic principles of how high-throughput and deep nucleotide sequencing data are generated, and how the massive number of reads will be interpreted and classified to obtained output formats for publications.

Proteomics further depends on the use and exploitation of big data from genomic and proteomic databases, their conversion to predicted in silico MS libraries and identification of proteins. The basic operation of data handling will be covered and practiced in the module (data format conversion, common formats). The principles will be taught, and students asked to solve practice data starting with single proteins moving to whole organisms. Examples for the use of proteomics from microbiology, plants and humans will be additionally presented and discussed. Aspects of quantitative proteomics methods will be introduced.

In the Metabolomics section mass spectral data will be applied to identify and quantify primary and secondary metabolites using mass spectrometry data. Targeted and untargeted metabolomic strategies will be introduced and the use of MS databases practiced. Basic aspects of compound quantification will be revisited (GLP, need for chromatographic separation, authentic standards, surrogate standards, calibration curves, reproducibility etc). NMR will be introduced as an alternative quantitative metabolomic technique with low sensitivity.

Metabolomics science further requires extensive use of statistical methods to compare large number of experimental data sets with one another each containing thousands of analytes. Hence the basic concepts of multivariate statistics (PCA, PLSDA, LDA, hierarchical clusters) and machine learning approaches will be introduced, and practice examples given to the students to allow metabolome comparison and biomarker identification. Examples of metabolomes will be discussed from microorganisms, plants and humans.

Recommended Knowledge

- Background literature on microbiology, molecular biology, protein biochemistry, protein structure and function.
- Background basic biochemistry literature on primary and secondary metabolites.
- Revision of mass spectrometry and chromatography.
- Basic knowledge on spectroscopic techniques (BSc level Analytical Methods), metabolism and proteins
- Understanding of chemical structure and bonding

Usability and Relationship to other Modules

According to Francis Crick's dogma of molecular biology (From genes to RNA to proteins to metabolites) this module is a complement to Omics I It has additional overlap with the elective structure elucidation module covering mass spectrometry. Statistical and quantitative methods discussed in other modules such Machine Learning, Quantitative

Experiments in Life Science and Computational Life Science complement the content in this module and bring it to life for students from a different perspective.

Intended Learning Outcomes

No	Competence	ILO
1	Appreciate	Appreciate information content of amplicon and metagenome sequencing as well as mass spectrometry data
2	Understand	Understand microbiomics and proteomics workflows and apply them for simple examples
3	Interpret	Interpret microbiome data to allow conclusions about the structure-function relationship in diverse microbial ecosystems
4	Use	Use proteomic databases in conjunction with proteomic LC-MS software to identify proteins and metabolites
5	Compare	Compare proteomes and quantitative proteomic data to design hypothesis on biological function and mechanisms
6	Compare	Compare metabolomes using multivariate statistical methods and conclude and speculate on their biological significance and identify biomarkers
7	Critically	Critically evaluate the reliability and significance of omics data, including sequencing and mass spectrometry results, in the context of biological research

Indicative Literature

- Microbiology, Bioanalytical Chemistry (Kurreck, Lottspeich, Engels, Springer 4th ed.)

Entry Requirements

Prerequisites	Guided Self-study
Co-requisites	None
Additional Remarks	None

Assessment and Completion

Components	Examination Type	Duration/Length	Weight (%)	Minimum	ILOs
Omics II - Microbiomics, Proteomics, and Metabolomics	Written Examination	120 minutes	100	45%	All

Module Achievements: None

4.16 Supramolecular Chemistry

Module Name	Supramolecular Chemistry
Module Code	2025-MQLS-CHE-01
Module ECTS	5
Study Semester	Mandatory status for: None Mandatory Elective status for: - 2025-AdMat-MSc 1 - 2025-QLS-MSc 1 - 2025-QLS-MSc 3 - 2025-AdMat-MSc 3
Duration	1 Semester
Program Affiliation	2025-QLS-MSc (Quantitative Life Science)
Module Coordinator(s)	Prof. Dr. Anna Tevyashova

Forms of Learning and Teaching	
Lecture	35
Exam Preparation	20
Independent Study	70
Workload Hours	125 hours

Module Components	Number	Type	CP
Supramolecular Chemistry	MQLS-CHE-01	Lecture	5

Module Description

The course aims to offer a comprehensive overview and foundational knowledge of supramolecular chemistry, focusing on non-covalent interactions. It covers a wide variety of these interactions, including hydrogen bonding, van der Waals forces, ionic interactions, and π - π stacking. Emphasis is placed on the principles governing these interactions and their roles in different chemical contexts. In drug design and discovery, understanding non-covalent interactions is essential for the rational design of drugs and inhibitors, which depend on specific binding to biological targets. In biomolecular engineering, the ability to manipulate and design supramolecular assemblies facilitates the creation of novel biomaterials and bio-inspired systems. Insights from supramolecular chemistry are also valuable in systems biology for modelling complex biological networks and understanding the emergent properties of biological systems. The principles of self-assembly and molecular recognition are fundamental in nanotechnology for developing nanoscale devices and materials. Furthermore, knowledge of non-covalent interactions enhances the development and application of advanced analytical techniques, such as spectroscopy and chromatography, which are crucial for probing molecular structures and dynamics. Supramolecular chemistry also provides the molecular framework for the quantitative methods essential for analyzing and modelling complex systems.

Recommended Knowledge

To prepare for this course, it is recommended that students have a solid background in basic chemistry, particularly in physical and organic chemistry. Reviewing foundational concepts in these areas will help students grasp the more advanced topics covered in the course. Additionally, reading introductory materials on supramolecular chemistry can provide a useful head start, ensuring that students are well-prepared to engage with the course content.

-Usability and Relationship to other Modules

The course plays a foundational role in understanding complex biological and chemical systems. Its focus on non-covalent interactions provides essential insights into the mechanisms underlying molecular recognition, self-assembly, and catalysts. These principles are critical for successful completion of Computational Life Science, Modelling and Simulation, Advanced Molecular Simulations, as well as Quantitative Experiments in Life Science modules. The principles of supramolecular chemistry are integral to the development of advanced materials.

Intended Learning Outcomes

No	Competence	ILO
1	Account	Account for fundamental concepts, methods and theories within supramolecular chemistry
2	Identify	Identify, analyze, and apply non-covalent interactions in diverse chemical contexts
3	Assess	Assess the current problems and research in the field
4	Anticipate	Anticipate the importance of supramolecular association within the life sciences and
5	Interpret	Interpret, analyze and evaluate experimental data of supramolecular interaction

Indicative Literature

- Jonathan W Steed, Jerry L Atwood, Supramolecular Chemistry, 2nd Ed Wiley-Blackwell, 2009, ISBN-13: 978-0470512340

Entry Requirements

Prerequisites	None
Co-requisites	None
Additional Remarks	None

Assessment and Completion

Components	Examination Type	Duration/Length	Weight (%)	Minimum	ILOs
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Supramolecular Chemistry	Written Examination	120 minutes	100	45%	All
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Module Achievements: None

4.17 Machine Learning in Cheminformatics

Module Name	Machine Learning in Cheminformatics
Module Code	2025-MQLS-COM-02
Module ECTS	5
Study Semester	Mandatory status for: None Mandatory Elective status for: - 2025-QLS-MSc 3
Duration	1 Semester
Program Affiliation	2025-QLS-MSc (Quantitative Life Science)
Module Coordinator(s)	Prof. Dr. Petr Popov

Forms of Learning and Teaching	
Lecture	17.5
Seminar	17.5
Independent Study	90
Workload Hours	125 hours

Module Components	Number	Type	CP
Machine Learning in Cheminformatics	MQLS-COM-02	Seminar	5

Module Description

Deep learning achieves remarkable results in many fields, including life sciences. This course is made for students that are ready to apply their skills to scientific applications, such as molecular design and drug discovery. Molecules play vital roles in our organism, constantly interacting with each other and performing all its functions.

Prediction of a biomolecule's properties, as well as design of molecules with target properties, are highly important yet unsolved problems and at the core of cheminformatics. The complex and rich nature of biomolecules can be represented as sequences, graphs, 3D objects, or high-dimensional descriptors, enabling the application of numerical methods in order to define their quantitative structure-activity relationship (QSAR).

The rapid accumulation of molecular data opened the gates for machine learning to be applied for such representations, and to derive powerful prediction models that outperform classical empirical methods. During this course students will be introduced to the open problems in cheminformatics and the state-of-the-art machine learning methods attempt to solve these problems. The course concludes with a final project aimed to solve cheminformatics problem of choice using machine learning.

Recommended Knowledge

Students will practice deep learning for drug discovery and molecular design problems. The course includes theoretical lectures, that cover basics of molecular structures, such that no prior knowledge of structural chemistry or biology is required. Seminars are python coding sessions, so python programming skills are required. Prior understanding of machine learning is an asset.

Usability and Relationship to other Modules

The course will be useful for computational chemists, computational biologists, data scientists as well as software engineers, who want to apply their skills for the cheminformatics.

Intended Learning Outcomes

No	Competence	ILO
1	Recall	Recall key concepts in cheminformatics
2	Explain	Explain the steps involved in the QSAR pipelines
3	Apply	Apply cheminformatics tools to analyze chemical compounds
4	Design	Design a machine learning-based QSAR pipeline

Indicative Literature

- Deep Learning for the Life Sciences, by Vijay Pande, Patrick Walters, Peter Eastman, Bharath Ramsundar Publisher: O'Reilly Media, Inc.
- Faulon, Jean-Loup, and Andreas Bender. Handbook of chemoinformatics algorithms. CRC press, 2010.

Entry Requirements

Prerequisites	Data Tools for the Life Sciences
Co-requisites	None
Additional Remarks	None

Assessment and Completion

Components	Examination Type	Duration/Length	Weight (%)	Minimum	ILOs
Machine Learning in Cheminformatics	Project Assessment		100	45%	All

Module Achievements: None

4.18 Intellectual Property and Commercialization

Module Name	Intellectual Property and Commercialization
Module Code	2025-MQLS-TS-03
Module ECTS	5
Study Semester	Mandatory status for: - 2025-QLS-MSc 3 - 2025-AdMat-MSc 3 Mandatory Elective status for: None
Duration	1 Semester
Program Affiliation	2025-QLS-MSc (Quantitative Life Science)
Module Coordinator(s)	

Forms of Learning and Teaching	
Lecture	35
Exam Preparation	20
Independent Study	70
Workload Hours	125 hours

Module Components	Number	Type	CP
Intellectual Property and Commercialization	MQLS-TS-03	Lecture	5

Module Description

Intellectual property (IP) refers to any form of creation of the mind, and its legal protection ensures that people earn recognition or financial benefit from what they invent or create. This module provides a critical examination of intellectual property (IP) legal aspects exploring the various categories of IP protection available for novel creations. The core focus is on copyright and patents, both granting exclusive rights to inventors for a defined period. We will analyze the criteria for patentability, the different patent types, and the process of patent acquisition. The module will delve into the advantages of patents (including the ability to control the production, sale, and import of inventions) but also explore the limitations of patents (e.g., territorial restrictions and eventual expiration). The module also teaches the various paths to commercialization of IP, including licensing, sale, and establishment of a startup, and it especially focuses on aspects of regulatory affairs, product registration, on the financing, private or public, of any of these options. Finally, it explains the construction of a business plan and a pitch deck, and the selection of the team for a startup. Interactions with, and visits to, initiatives at the University, in the Constructor ecosystem, and in the city-state of Bremen are anticipated to provide an integrated view and help with first steps. By the end, participants will gain a comprehensive understanding of how copyrights and patents can be leveraged to protect inventions and drive innovation. This module

is particularly valuable for researchers, aspiring inventors, and those with an entrepreneurial spirit.

Recommended Knowledge

The previous modules in the Transferable Skills track, i.e., Scientific Presentations and Scientific Writing, are recommended preparation, while not being strict prerequisites.

Usability and Relationship to other Modules

The module belongs to the Transferable Skills.

Intended Learning Outcomes

No	Competence	ILO
<i>A. Understand Intellectual Property</i>		
1	Master	Master Intellectual Property: Comprehend IP concepts, distinguish IP categories, and evaluate their protection mechanisms
2	Copyright:	Copyright: Grasp legal concepts of copyright
3	Patent	Patent Expertise: Grasp patent law, including patentability criteria, types, acquisition, enforcement, and impact on innovation
<i>B. Commercialize Intellectual Property</i>		
4	IP	IP Commercialization: Explore diverse paths for IP commercialization, identify funding sources, and construct effective business plans
5	Entrepreneurial	Entrepreneurial Mindset: Develop entrepreneurial skills, build high-performing teams, and leverage resources for IP venture success
<i>C. Achieve Strategic IP Management</i>		
6	IP	IP Strategy: Analyze real-world case studies to understand IP's role in business strategy, risk management, and competitive advantage
7	IP	IP Valuation and Licensing: Evaluate IP assets, negotiate licenses, and understand the complexities of IP transactions
8	IP	IP and Society: Examine the ethical, legal, and societal implications of IP, considering issues like open innovation and IP policy

Indicative Literature

- Reingand, N. (ed.), Intellectual Property in Academia: A Practical Guide for Scientists and Engineers, CRC Press, 2011, ISBN-10 1439837007

Entry Requirements

Prerequisites	None
Co-requisites	None
Additional Remarks	None

Assessment and Completion

Components	Examination Type	Duration/ Length	Weight (%)	Minimum	ILOs
Intellectual Property and Commercialization	Oral Examination	30 minutes	100	45%	All

Module Achievements: None

4.19 Master Thesis QLS

Module Name	Master Thesis QLS
Module Code	2025-MQLS-THE-01
Module ECTS	30
Study Semester	Mandatory status for: - 2025-QLS-MSc 4 Mandatory Elective status for: None
Duration	1 Semester
Program Affiliation	2025-QLS-MSc (Quantitative Life Science)
Module Coordinator(s)	

Forms of Learning and Teaching	
Laboratory	560
Exam Preparation	20
Independent Study	170
Workload Hours	750 hours

Module Components	Number	Type	CP
Master Thesis QLS	MQLS-THE-01	Thesis	30

Module Description

The aim of this module is to train students to motivate, design, carry out and document a research project in one of the research groups of the faculty of Quantitative Life Science. Some familiarity with the requisite experimental techniques and analytical skills will typically have been acquired in one of the preceding Lab Rotations (I, II and III). The thesis topic is determined in mutual agreement with the laboratory head. They may arise from the ongoing research in the instructor's own research group, but it is also possible for a student to adopt a topic of his/her own choice provided the instructor agrees to supervise it. The project selected should be a topical piece of novel science with a view of being published in a peer-reviewed scientific journal. The thesis work comprises the full cycle of a scientific research endeavor: (i) identifying a relevant open research question, (ii) carrying out a literature survey to put the planned work in its context and relate it to the state of the art (SoA), (iii) formulate a concrete research objective, (iv) design a research plan including a statement of criteria to evaluate the success of the project, (v) carry out the plan (with the possibility to change the original plan when necessary), (vi) document the results, (vii) analyze the results with respect to the SoA, the original objective, and the success criteria, and (viii) and summarize all of this work in the thesis document. Moreover, the student is expected to actively participate in the meetings of and engage with the respective research group. All of this work should be done with as much self-guidance as can be reasonably expected. The laboratory head will likely give substantial guidance for (i), (iii) and (iv), whereas the other aspects will be addressed with larger degrees of self-guidance and with the support of the research group.

Practical laboratory and instrument training will be provided by members of the relevant research groups (Postdocs, technicians and experienced PhD students). A research proposal document summarizing (i) – (iv) is expected as an interim result and milestone (target size: 5 pages). At the end of the course, there will be a conference-style meeting where all students present their work.

The project concludes with a thesis, and an oral defense thereof.

Recommended Knowledge

- Study indicative research literature and methods in the selected research group.
- Proficiency in the area of the chosen thesis topic.

Usability and Relationship to other Modules

The master thesis can build on the Lab Rotation I, II or III, but students are free to choose a different topic and a different supervisor for the master thesis.

Intended Learning Outcomes

No	Competence	ILO
1	Discuss	Discuss, at a professional level, a circumscribed segment of the hosting group's research area
2	Compare	Compare and select experimental methods, as required for the project, at a professional level
3	Design	Design and implement the full cycle of a novel scientific research project in a professional manner
4	Formulate	Formulate a research proposal such that that it could serve as a funding proposal
5	Author	Author a Master Thesis at the level expected from a scientific publication
6	Examine	Examine and defend his own research in the wider scientific context
7	Demonstrate	Demonstrate general professional skills like communication, presentation and time management

Indicative Literature

Entry Requirements

Prerequisites	Laboratory Rotation QLS I Laboratory Rotation QLS II Laboratory Rotation QLS III
Co-requisites	None
Additional Remarks	

Assessment and Completion

Components	Examination Type	Duration/ Length	Weight (%)	Minimum	ILOs
Master Thesis QLS	Thesis	30-60 pages	80	45%	All
	Oral Examination	30	20		All

Module Achievements: None.

5.1 Intended Learning Outcome Matrix MSc Quantitative Life Science

MSc Quantitative Life Science				Guided self-study	Data Tools for Life Sciences	Experimental Techniques	Omics I - Genomics and Transcriptomics	Computational Life Sciences - Modelling and Simulations	Omics II - Microbiomics and Metabolomics	Molecular Simulations	Machine Learning in Cheminformatics	Supramolecular Chemistry	Structure Elucidation	Biochemical Engineering: From Cells to Processes	Quantitative Cell Biology	Lab Rotation QLS I	Lab Rotation QLS II	Lab Rotation QLS III	Scientific Presentations	Scientific Writing	Intellectual Property and Commercialization	Master Thesis
Semester				1	1	2	2	3	3	3	3	1 or 3	2	1 or 3	4	1	2	3	1	2	3	4
Mandatory/ Mandatory elective				m	m	m	m	m	m	me	me	me	me	me	me	m	m	m	m	m	m	m
ECTS Credits				5	5	5	5	5	5	5	5	5	5	5	5	10	10	10	5	5	5	30
Competencies*																						
Program Learning Outcomes	A	E	P	S																		
Describe complex technical and conceptual advances in the Life Sciences in depth and with specific focus on omics techniques, like genomics, transcriptomics, proteomics, and metabolomics.	x						x	x	x	x	x	x	x	x	x				x	x		
Understand common computational tools, algorithms, and databases for data processing, analysis and visualization used in the life sciences.	x					x	x	x	(x)	x			(x)	(x)								
Critically examine and creatively apply intricate techniques to perform quantitative experiments, obtain reliable data, and answer complex scientific questions in the life sciences at high scientific and ethical standards.	x				x		x	x	x			x	x	x	x	x	x	x	x			x
Scrutinize and implement innovative bioinformatics approaches to obtain, examine, and compare Life Science data.	x					x		x	x	x	x			x		(x)	(x)	(x)	x			
Construct comprehensive mathematical models and detailed computational simulations to generate and test hypothesis in the field of biology or chemistry.	x				x			x		x	x					(x)	(x)	(x)				
Scrutinize and critique scientific results, experimental approaches, and research hypothesis from all fields of quantitative life science.	x					x	x	x	x	x									x			x
Construct original and innovative research hypothesis and design experiments to test them individually or as a team in actual research projects and a master thesis.	x	x		x			x		x	x	x	x	x	x	x				x			x
Apply professional writing, communication, and presentation techniques, combined with state-of-the-art visualization tools to effectively communicate with scientific and non-scientific audiences.	x	x		x															x	x	x	
Understand the chances, risks, and approaches of applying scientific research in modern society considering ethical and commercial aspects while following and defending ethical, scientific, and professional standards.		x	x	x																x	x	
Take responsibility for their own learning, personal and professional development and role in diverse teams, evaluating critical feedback and self-analysis.		x	x	x	x																	x
Assessment Type																						
Written examination					x	x	x	x	x	x		x	x	x	x							
Term paper																				x		
Essay																						
Project report						x																
Poster presentation										x							x	x	x			
Laboratory report																						
Program Code																						
Oral examination																					x	x
Presentation															x		x	x	x			
Practical Assessments																						
Project Assessments											x											
Portfolio Assessments																						
Master Thesis																						x
Module Achievements							x															

*Competencies: A-scientific/academic proficiency; E-competence for qualified employment; P-development of personality; S-competence for engagement in society